



Revue rapide version 1 : Que sait-on du temps que peut survivre le virus, avec un potentiel d'infection, sur des surfaces que l'on trouve dans la collectivité?

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Veillez noter : Cette revue a peut-être été mise à jour. Consultez la version la plus récente de cette revue en visitant le Service rapide de données probantes sur la COVID-19 du Centre de collaboration nationale des méthodes et outils, au lien ci-dessus.

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Cette mise à jour a été réalisée avec la contribution de collègues du Service rapide de données probantes sur la COVID-19 de Public Health England.

Cette revue rapide est destinée à des fins d'information générale seulement. Les renseignements qui figurent dans le présent revue rapide sont fournis « en l'état » et l'Université McMaster ne fait aucune garantie, promesse et/ou représentation de quelque nature que ce soit, expresse ou implicite, quant à la nature, la norme, l'exactitude, l'exhaustivité, la fiabilité ou autre des renseignements fournis dans le présent revue rapide, ni quant à la pertinence ou autre des renseignements par rapport à des circonstances particulières. L'Université McMaster n'accepte aucune responsabilité quant à l'exactitude, au contenu, à l'exhaustivité, à la légalité, à la fiabilité ou à l'utilisation des renseignements contenus dans le présent revue rapide.

Les auteures déclarent n'avoir aucun conflit d'intérêts à divulguer.

Résumé

Contexte

Alors que la transmission communautaire de la maladie à coronavirus 2019 (COVID-19) se poursuit à travers le monde, il est important de comprendre le rôle que peut jouer la transmission indirecte par les surfaces dans la collectivité.

Cette revue rapide a été produite pour soutenir la réponse de l'Agence de la santé publique du Canada à la pandémie de coronavirus 2019 (COVID-19). Cette revue vise à recenser, évaluer et résumer les nouvelles données de recherche à l'appui de la prise de décision fondée sur des données probantes.

Cette revue rapide inclut les données probantes disponibles au 31 décembre 2020 pour répondre à la question suivante : **Que sait-on du temps que peut survivre le virus, avec un potentiel d'infection, sur des surfaces que l'on trouve dans la collectivité?**

Qu'est-ce qui a changé dans cette version?

- Depuis la dernière version de cette revue (31 juillet 2020), une quantité considérable de données probantes se sont ajoutées. Pour pouvoir réaliser la mise à jour de cette revue rapide, nous avons affiné la question de recherche afin qu'elle ne porte que sur les surfaces se trouvant dans la collectivité (et non en milieu hospitalier ou clinique). Pour cette raison, 18 études qui avaient été incluses dans la revue originale ont été exclues de cette mise à jour. Une liste des références exclues peut être consultée [ici](#).
- Cette mise à jour inclut quatre nouvelles synthèses, la mise à jour d'une synthèse précédemment incluse, et 22 nouvelles études uniques.

Points clés

- Dans plusieurs synthèses et études transversales, des données probantes cohérentes indiquent que des fragments du SRAS-CoV-2 peuvent être détectés sur des surfaces dans la collectivité pendant une période pouvant aller jusqu'à sept jours. Le degré de certitude des données probantes est considéré comme modéré (GRADE), et les conclusions pourraient changer à mesure que de nouvelles informations apparaîtront. Cependant, la plupart de ces études mesurent le matériel génétique viral, et elles ne distinguent donc pas les virus vivants des virus morts ou des fragments de virus. Une seule étude mesure les virus viables (qui ont un potentiel d'infection) dans les échantillons, et elle n'en a trouvé aucun.
- Globalement, des fragments viraux peuvent être détectés sur des surfaces, mais ces fragments ne sont peut-être pas viables. Le degré de certitude de ces données probantes est considéré comme faible (GRADE) et les conclusions pourraient changer à mesure que de nouvelles informations apparaîtront.
- Dans les études qui indiquent si un nettoyage des surfaces avait été fait avant l'échantillonnage, les procédures de désinfection ou de nettoyage ont invariablement diminué ou éliminé la détection de fragments de SRAS-CoV-2. Le degré de certitude des données probantes est modéré (GRADE), et les conclusions pourraient changer à mesure que de nouvelles informations apparaîtront.

- Les résultats d'études en laboratoire indiquent que le SRAS-CoV-2 peut rester viable plus longtemps sur des surfaces plus lisses, comme le plastique ou l'acier, que sur du carton ou du coton. Toutefois, ces études ont souvent des concentrations de départ beaucoup plus élevées que celles que l'on trouve dans l'environnement. Les durées rapportées varient beaucoup, mais les données indiquent une plus grande stabilité à des températures plus basses (comme 4 °C) et une dégradation plus rapide à mesure que les températures augmentent. La qualité des études ne peut pas être évaluée avec l'outil GRADE, car ces données probantes sont seulement tirées d'études en laboratoire et qu'on ignore l'applicabilité de ces résultats au monde réel.

Aperçu des données probantes et lacunes dans les connaissances

- Plusieurs études ayant recueilli des échantillons de surface dans la collectivité ont détecté des fragments de SRAS-CoV-2 par la RT-PCR en temps réel dans des échantillons tirés de diverses surfaces, en particulier les surfaces dans les chambres à coucher et les salles de bain et les surfaces fréquemment touchées, comme les poignées de porte.
- Conformément aux résultats d'autres synthèses, la probabilité de trouver des fragments de SRAS-CoV-2 sur des surfaces échantillonnées dans la collectivité varie selon les études. Il n'est pas possible de déterminer dans quelle mesure cette variabilité est due aux différentes méthodes d'échantillonnage utilisées, lesquelles influencent la probabilité de détecter des particules du virus. Par exemple, si certaines études ont fait un échantillonnage aléatoire de surfaces, la plupart se sont intéressées aux surfaces les plus touchées afin de maximiser les probabilités de détection.
- Bien que des particules de virus aient été détectées, les méthodes de dépistage employées dans plusieurs de ces études ne sont pas aussi rigoureuses que la norme actuelle (laquelle exige que trois ARN cibles soient amplifiés par RT-PCR en temps réel [Corman *et al.*, 2020]), car elles n'ont utilisé qu'un ou deux ARN cibles. Selon Moreno *et al.*, la plupart des échantillons ayant obtenu un résultat positif n'ont eu un résultat positif que pour l'un des trois ARN cibles. De leur côté, aucun des échantillons de Di Carlo *et al.* ne contenait au moins deux cibles ayant obtenu un résultat positif. La plupart des études ne font pas de distinction entre les virus vivants et les virus morts ou les fragments de virus.
- Une seule étude transversale, par Döhla *et al.*, a tenté de détecter des virus viables (des virus vivants qui ont conservé un potentiel d'infection) par test de culture virale. Elle n'en a détecté aucun, malgré qu'elle ait trouvé des échantillons ayant obtenu un résultat positif par RT-PCR en temps réel (ce qui laisse entendre que ces échantillons ayant obtenu un résultat positif ne contenaient peut-être pas de virus viables). Ainsi, il semble que le fait de toucher une surface contaminée entraîne un faible risque d'infection, même si les données probantes sont très limitées, puisqu'une seule étude a employé un test de culture virale cherchant à détecter des virus vivants.
- Plusieurs procédures de désinfection ou de nettoyage ont été effectuées après la contamination, mais avant l'échantillonnage, et celles-ci réduisent ou éliminent invariablement les échantillons ayant obtenu un résultat positif à un test RT-PCR en temps réel de dépistage du SRAS-CoV-2. Sur les huit études qui mentionnent explicitement que les surfaces avaient été nettoyées avant la collecte d'échantillons, cinq n'ont trouvé que des échantillons négatifs et trois, un petit nombre d'échantillons

positifs. La plupart des études présentent des pratiques de nettoyage variées ou ne sont pas explicites au sujet de celles-ci. Toutefois, la présence d'une procédure de nettoyage réduit systématiquement la détection de particules virales.

- Dans des conditions de laboratoire contrôlées, dont certaines utilisaient des titres initiaux élevés, la période au cours de laquelle les virus viables pouvaient être détectés (par des tests de culture virale) sur des surfaces non poreuses (dures) variait de 3 à 28 jours. Sur des surfaces poreuses comme du tissu, les études qui ont fait des tests sur plusieurs points dans le temps révèlent que la majorité étaient absorbés rapidement après l'application, mais qu'ils pouvaient toujours être détectés après une période allant de trois heures à 14 jours. Les études en laboratoires utilisent souvent des concentrations de départ qui sont considérablement plus élevées que celles que l'on observe dans l'environnement. Ces concentrations de départ influenceront les durées de survie, ce qui rend difficile la généralisation des résultats d'études en laboratoire aux contextes réels.
- Sept études en laboratoire ont étudié l'effet de la température sur la survie d'un virus du SRAS-CoV-2 viable. Des températures allant de 4 °C à 70 °C ont fait l'objet de tests. Six études ont révélé une plus grande stabilité à des températures plus faibles, indiquant une dégradation plus rapide à mesure que les températures augmentent. Une étude a démontré la plus grande stabilité à 30 °C, le virus étant le plus stable à la température ambiante.
- Une étude a examiné l'effet de l'humidité relative sur la survie du virus. Elle a révélé que la dégradation la plus rapide se produisait à une humidité relative de 65 %, tandis qu'une dégradation plus lente se produisait à une humidité relative plus faible (40 %) et plus élevée (85 %). Toutefois, l'applicabilité de ces résultats à des contextes réels est inconnue.

Méthodologie

Question de recherche :

Que sait-on du temps que peut survivre le virus, avec un potentiel d'infection, sur des surfaces que l'on trouve dans la collectivité?

Recherche

Les bases de données suivantes ont été fouillées le 4 janvier 2021 en utilisant les termes clés "indirect transmission", "fomite", "surface", and "touch" avec une limite au 31 décembre 2020 :

- [MEDLINE](#)
- [Embase](#)
- [Trip Medical Database](#)
- [COVID-19 Evidence Alerts](#) from McMaster PLUS™
- [Public Health +](#)
- [COVID-19 Living Overview of the Evidence \(L·OVE\)](#)
- [McMaster Health Forum](#)
- [Prospero Registry of Systematic Reviews](#)
- [MedRxiv preprint server](#)
- NCCMT [COVID-19 Rapid Evidence Reviews](#)
- NCCDH [Equity-informed Responses to COVID-19](#)
- NCCEH [Environmental Health Resources for the COVID-19 Pandemic](#)
- NCCID [Disease Debrief](#)
- NCCIH [Updates on COVID-19](#)
- NCCHPP [Public Health Ethics and COVID-19](#)
- [Uncover \(USHER Network for COVID-19 Evidence Reviews\)](#)
- Centers for Disease Control and Prevention's [Morbidity and Mortality Weekly Report \(MMRW\)](#)
- [Institute national d'excellence en santé et en services sociaux \(INESSS\)](#)
- [BC Centre for Disease Control \(BCCDC\)](#)
- [Public Health Ontario](#)
- [Institut national de santé publique du Québec \(INSPQ\)](#)
- Alberta Health Service's [COVID-19 Resources for AHS Staff & Health Professionals](#)

Une copie de la stratégie de recherche complète peut être consultée à [lien](#).

Qu'est-ce qui a changé dans cette version en ce qui a trait aux méthodes?

La littérature sur ce sujet a pris beaucoup d'ampleur depuis la dernière version de cette revue. Pour réaliser une synthèse rapide et ciblée des données probantes, les critères d'inclusion et d'exclusion ont été affinés afin d'exclure les études réalisées en milieu clinique ou hospitalier. Plusieurs études qui avaient été incluses dans les versions antérieures de cette revue en ont maintenant été exclues. La liste des références exclues peut être obtenue en cliquant sur ce [lien](#). Cette revue présente une synthèse des données probantes sur la transmission du virus qui cause la COVID-19 sur des surfaces que l'on trouve habituellement dans la collectivité.

Critères de sélection des études

Les résultats de la recherche ont d'abord été examinés pour recenser les directives et les synthèses récentes. Les études uniques ont été incluses si aucune synthèse n'était disponible ou si des études uniques ont été publiées après que la recherche ait été effectuée à partir de la synthèse. Les sources de langue anglaise évaluées par les pairs et les sources publiées avant l'impression et avant l'évaluation par les pairs ont également été incluses. Les sources de surveillance ont été exclues. Lorsqu'ils sont disponibles, les conclusions des synthèses et les guides de pratique clinique sont présentés en premier, car ils tiennent compte de l'ensemble des preuves disponibles et peuvent donc être appliqués largement aux populations et aux milieux.

| | Critères d'inclusion | Critères d'exclusion |
|--------------|--|--|
| Population | Surfaces inanimées | Milieux cliniques Milieux hospitaliers |
| Intervention | Exposition au SRAS-CoV-2 | Exposition par contact étroit avec une personne infectée |
| Comparaison | - | Études en laboratoire dont le principal objectif était de comparer différents produits de désinfection/nettoyage |
| Résultats | Détection du virus du SRAS-CoV-2 ou ARN Infection à la COVID-19 | |

Extraction et synthèse des données

Pour les synthèses, les données relatives à la conception de l'étude, au cadre, à l'emplacement, aux caractéristiques de la population, aux interventions ou à l'exposition et aux résultats ont été extraites lorsqu'elles étaient déclarées.

Évaluation de la qualité des données probantes

Nous avons évalué la qualité des données probantes incluses en utilisant des outils d'évaluation critique, comme nous le décrivons ci-dessous. L'évaluation de la qualité a été réalisée par un examinateur et vérifiée par un deuxième examinateur. Les conflits ont été résolus par la discussion. Pour certaines des données probantes incluses, aucun outil approprié n'a été trouvé, ou l'équipe de revue n'avait pas l'expertise nécessaire pour évaluer leur qualité méthodologique. Les études pour lesquelles aucune évaluation de la qualité n'a été effectuée sont indiquées dans les tableaux de données.

Méthodologie de l'étude Outils d'évaluation critique

Synthèse Assessing the Methodological Quality of Systematic Reviews (AMSTAR)
[AMSTAR 1 Tool](#)

Prévalence Joanna Briggs Institute (JBI) [Checklist for Prevalence Studies](#)

Les évaluations de la qualité effectuées pour chaque étude incluse sont disponibles sur demande.

L'approche [GRADE](#) (Grading of Recommendations, Assessment, Development and Evaluations) a été utilisée pour évaluer la certitude des résultats sur la base de huit domaines clés.

Selon l'approche GRADE en matière de qualité des données probantes, les **études observationnelles**, telles que celles incluses dans cette revue, fournissent des données probantes de **faible qualité**. Cette évaluation peut être réduite encore davantage en fonction d'autres domaines :

- un risque de biais élevé;
- l'incohérence des effets;
- le caractère indirect des interventions/résultats;
- des imprécisions dans l'estimation de l'effet;
- un biais de publication.

À l'inverse, elle peut être rehaussée sur la base des domaines suivants :

- un effet important;
- une relation dose-effet;
- une prise en compte des variables confusionnelles.

Pour chaque résultat, la certitude globale des données probantes a été déterminée en tenant compte des caractéristiques des données probantes dont on dispose (des études observationnelles, dont certaines n'ont pas été évaluées par les pairs, des variables confusionnelles potentielles qui n'ont pas été prises en compte, des essais et des protocoles d'essais différents, et une absence de groupes de comparaison valides). Un jugement selon lequel « la certitude globale est très faible » signifie que les résultats risquent fort de changer à mesure que de nouvelles données probantes apparaissent.

Résultats

Synthèse de la qualité des données probantes

Dans cette mise à jour, 4 nouvelles synthèses, une mise à jour à une synthèse précédemment incluse, 22 nouvelles études individuelles ont été recensées, et 18 études ont été considérées inadmissibles, pour un total de 41 publications portant sur la question de recherche. La qualité des données probantes incluses dans cette revue se décrit comme suit :

| Question(s) de recherche | Données probantes incluses | | Certitude globale des données probantes |
|--|----------------------------|----|---|
| Que sait-on du temps que peut survivre le virus, avec un potentiel d'infection, sur des surfaces que l'on trouve dans la collectivité? | Synthèses terminées | 7 | Faible-moderée |
| | Synthèses en cours | 2 | |
| | Études individuelles | 32 | |

Attention

Comme il faut rendre rapidement disponibles les nouvelles données probantes sur la COVID-19, plusieurs études émergentes n'ont pas été révisées par des pairs. Pour cette raison, nous vous conseillons la prudence quand vous utilisez et interprétez les données probantes incluses dans cette revue rapide. Nous avons fourni une synthèse de la certitude globale des données probantes afin de soutenir le processus de prise de décision. Lorsque c'est possible, nous vous recommandons de fonder vos décisions sur les données probantes de la plus haute qualité possible.

Il est important de noter que nous n'avons pas évalué la qualité méthodologique des études en laboratoire. En raison de la nature hautement technique de ces études, nous recommandons vivement de consulter un expert en matière de contenu pour éclairer la prise de décision.

Tableau 1 : Synthèses

| Reference | Date Released | Description of Included Studies | Summary of Findings | Quality Rating: Synthesis | Quality Rating: Included Studies |
|---|--|--|---|---------------------------|----------------------------------|
| New evidence reported March 5, 2021 | | | | | |
| Bedrosian, N., Mitchell, E., Rohm, E., Rothe, M., Kelly, C., String, G., & Lantagne, D. (2020). A systematic review of surface contamination, stability, and disinfection data on SARS-CoV-2 (through July 10, 2020) . <i>Environmental Science and Technology</i> . Epub ahead of print. | Nov 23, 2020 (Search completed Jul 10, 2020) | This review included: <ul style="list-style-type: none"> • 35 studies on surface contamination • 16 studies on surface stability • 27 studies on surface disinfection | <p>Surfaces contamination in the community: 2.5% of household surfaces positive to SARS-CoV-2; 14% in non-household accommodation; and 14% in outdoor settings (including 25% high-touch surfaces and 23% hard furniture).</p> <p>SARS-CoV-2 half-life: 2.3-17.9 hours on stainless steel; 2.3-15.3 hours on plastic; 2.3-15.3 hours on nitrile. Half-life decreases as temperature and humidity increase.</p> <p>A 99.9% virus reduction can be obtained with sunlight, ultraviolet light, ethanol, hydrogen peroxide and hypochlorite.</p> <p>Knowledge gap on the contribution of fomite to SARS-CoV-2 transmission.</p> | Low | Not reported |
| Bueckert, M., Gupta, R., Gupta, A., Garg, M., & Mazumder, A. (2020). Infectivity of SARS-CoV-2 and other coronaviruses on dry surfaces: Potential for indirect transmission . <i>Materials</i> , 13(22), 5211. | Nov 18, 2020 (Search date not reported) | This review included 26 studies, of which 15 were related to SARS-CoV-2. Probably all laboratory-based studies, although not clearly stated | <p>Overall, porous substrates seem to inactivate SARS-CoV-2 faster than non-porous material (with some exceptions, such as N-95 and surgical masks on which SARS-CoV-2 appear to be remarkably stable).</p> <p>Correlation between material wettability and SARS-CoV-2 stability.</p> <p>Cotton and cellulose-based materials usually attenuate SARS-CoV-2 quicker than other materials (besides copper).</p> <p>SARS-CoV-2 persistence inversely related to temperature and humidity, and sunlight might inactivate the virus.</p> | Low | Not reported |

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|---|---|--|---|--|---------------------|
| <p>Fernández-Raga, M., Diaz-Marugan, L., Garcia Escolano, M., Bort, C., & Fanjul, V. (2020). SARS-CoV-2 viability under different meteorological conditions, surfaces, fluids and transmission between animals. <i>Environmental Research</i>, 192, 110293.</p> | <p>Oct 2, 2020 (Search completed Jun 29, 2020)</p> | <p>Approximately 6 studies were included related to SARS-CoV-2, although this is not clear</p> | <p>SARS-CoV-2 can stay for a variable period on different surfaces (from hours to days) maintaining its infective potential. However, touching a contaminated surface is only infective if contact with the surface ends up in mucosal membranes.</p> | <p>Low</p> | <p>Not reported</p> |
| <p>Meyerowitz, E. A., Richterman, A., Gandhi, R. T., & Sax, P. E. (2020). Transmission of SARS-CoV-2: A review of viral, host, and environmental factors. <i>Annals of Internal Medicine</i>, 174(1), 69-79.</p> | <p>Sep 17, 2020 (Search completed Sep 7, 2020)</p> | <p>This review included:</p> <ul style="list-style-type: none"> • 1 laboratory-based study • 18 studies sampling contaminated environmental surfaces | <p>There is no conclusive evidence for fomite or direct contact transmission of SARS-CoV-2 in humans, as reports suggesting fomite transmission are circumstantial (e.g., infected persons reporting no direct contact with a case, suggesting transmission via shared common facilities).</p> <p>Authors suspect the levels of viral RNA or live virus remaining on surfaces are unlikely to cause infection.</p> | <p>Low</p> | <p>Not reported</p> |
| <p>Usher Institute. (2020, Aug 15). Summary: What is the evidence for indoor transmission of SARS-CoV-2?</p> | <p>Aug 15, 2020 (Search completed May 21, 2020)</p> | <p>This review included 66 studies (to answer a range of questions). Number of studies relating to surfaces or types of study not reported</p> | <p>Laboratory-based studies suggest the virus may persist longer on smooth surfaces, such as plastic or stainless steel (up to 72 or 48 hours, respectively), than cardboard (up to 24 hours); and at low temperatures (highly stable at 4°C, not at 70°C) and damp conditions.</p> <p>Although viral RNA was detected on a range of objects in clinical and non-clinical settings, 3 studies which quantified the amount of virus present found minimal amounts.</p> <p>One epidemiological study reported fomite transmission, through occupying the same seat as the index case in a church.</p> | <p>Low</p> <p>NOT PEER REVIEWED</p> | <p>Low</p> |

| Previously reported evidence | | | | | |
|--|--|---|--|-------------------------------------|--------------|
| Fiorillo, L., Cervino, G., Matarese, M., D'Amico, C., Surace, G., Paduano, V., ... Cicciù, M. (2020). COVID-19 Surface Persistence: A Recent Data Summary and Its Importance for Medical and Dental Settings . <i>International Journal of Environmental Research and Public Health</i> , 17(9), 3132. | Apr 30, 2020 (Search date not reported) | This review included 4 laboratory-based studies, only one focused on SARS-CoV-2 (also included above). | SARS-CoV-2 persisted longest on plastic and stainless steel. The virus was not detectable after 4 hours on copper and 24 hours on cardboard. These findings are consistent with other coronaviruses. | Moderate | Not reported |
| National Academies of Sciences, Engineering, and Medicine. (2020, Mar 27). Rapid expert consultation update on SARS-CoV-2 surface stability and incubation for the COVID-19 pandemic . | Mar 27, 2020 (Search date not reported) | This review included: <ul style="list-style-type: none"> • Experimental laboratory-based studies • Prevalence studies | <p>Two lab-based studies were described, as well as preliminary results from in-progress studies via personal communication.</p> <p>SARS-CoV-2 showed greater stability on smooth surfaces (glass, banknote, stainless steel, plastic); no infectious virus was detected after 4-7 days. Also, infectious virus was detectable on the outer layer of a surgical mask after 7 days.</p> <p>Across prevalence studies, there were variable rates of surface samples testing positive for SARS-CoV-2 primarily using reverse transcriptase-polymerase chain reaction (RT-PCR) for testing.</p> <p>Samples across studies were collected both prior to and after cleaning/ disinfection, from different sites (personal rooms, common areas) and across different settings (hospitals, cruise ship).</p> | Low NOT PEER REVIEWED | Not reported |

Tableau 2 : Synthèses en cours

| Title | Anticipated Release Date | Setting | Description of Document |
|--|--------------------------|---------------|--|
| Previously reported evidence | | | |
| Dalla Nora, V., Azevedo, N. & Rosa, D. (2020). <i>Survival of SARS-CoV-2 on different surfaces of the dental office and the effective disinfection agents.</i> PROSPERO, CRD42020188152. | Oct 15, 2020 | Dental office | This systematic review aims to explore the survival time of SARS-CoV-2 on different surfaces in dental offices and determine decontamination agents and concentration levels for effective disinfection. |
| Deliga Schroder, A. G., Guariza Filho, O., Neto, J. S., Gonçalves, F. M., Bittencourt Basso, I., Sampaio Santos, R., ... Nogueira Cortz Ravazzi, G. M. (2020). <i>COVID-19 survival time on inanimate surfaces: a systematic review.</i> PROSPERO, CRD42020185643. | Jun 30, 2020 | Multiple | This systematic review aims to explore survival time of SARS-CoV-2 on different types of inanimate surfaces. |

Tableau 3 : Études individuelles

| Reference | Date Released | Study Design | Setting | Method of testing and timing | Summary of findings | Quality Rating: |
|---|---------------|--------------|---|--|---|-----------------------------------|
| Prevalence Studies | | | | | | |
| New evidence reported March 5, 2021 | | | | | | |
| Ming, Z., Han, S., Deng, K., Ha, Y., Kim, S., Reyes, E., ... Samadpour, M. (2020). Environmental monitoring shows SARS-CoV-2 contamination of surfaces in food plants. <i>Preprint.</i> | Dec 11, 2020 | Prevalence | Surface samples collected from 116 food production facilities. USA | RT-PCR Samples were collected by plant personnel in different areas and on different surface types between Mar 17 and Sep 3, 2020. 5 facilities followed longitudinally for timeline data analyses in function of preventive measures (but no details provided on these measures). No information on cleaning protocols provided. | 278/22,643 (1.23%) of samples tested positive for SARS-CoV-2, and 62/116 (53%) of facilities had at least 1 positive sample. Virus commonly found on frequently touched surfaces such as doorknobs/handles, tables, computer devices and sanitizer dispensers. Receiving rooms and entrance had the highest occurrence of positive samples. In 3/5 facilities, decreasing trend of daily positive rate observed. | Low <i>PREPRINT</i> |

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| <p>Moreno, T., Pintó, R. M., Bosch, A., Moreno, N., Alastuey, A., Minguillón, M. C., ... Querol, X. (2021). Tracing surface and airborne SARS-CoV-2 RNA inside public buses and subway trains. <i>Environment International</i>, 147, 106326. Epub ahead of print.</p> | <p>Dec 9, 2020</p> | <p>Prevalence</p> | <p>Surface and air samples collected from buses and subway trains.</p> <p>Spain</p> | <p>RT-PCR</p> <p>75 samples from buses and 24 from trains samples collected between May and Jul 2020 (78 surface samples, 12 air samples, 9 samples from air-conditioning filters).</p> <p>Bus samples were collected before and other after cleaning (bleach or ozone); train samples were collected at the end of the day.</p> <p>82 of the 99 samples were analysed.</p> | <p>30/82 were positive for SARS-CoV-2.</p> <p>Train: 6/15 surface samples positive 2/6 air samples positive 0/3 filters samples positive</p> <p>Bus: 13/30 surface samples positive; 3 of these 13 surfaces were still positive after cleaning 1/6 air samples positive 3/6 filters samples positive</p> <p>Most of the positive samples (24/30) tested positive for only 1 of the 3 RNA targets; risk of infections considered to be extremely low.</p> | <p>Moderate</p> |
| <p>Maestre, J. P., Jarma, D., Yu, C., Siegel, J., Horner, S., & Kinney, K. A. (2020). Distribution of SARS-CoV-2 RNA signal in a home with COVID-19 positive occupants. <i>Preprint</i>.</p> | <p>Dec 2, 2020</p> | <p>Prevalence</p> | <p>Surface and dust samples collected from a household with 2 confirmed COVID-19 cases 1 month after symptom resolution (2 months after symptom onset)</p> <p>USA</p> | <p>RT-PCR</p> <p>22 surface samples collected by household member using a sampling kit provided by the research team.</p> <p>Dust samples collected from bedroom and HVAC filter using a handheld vacuum cleaner.</p> <p>Information on cleaning practices and surface material obtained by remote survey.</p> | <p>11/24 (46%) of surface samples were positive for SARS-CoV-2, mostly from the bedroom, surface of HVAC filter, and living room.</p> <p>Samples from surfaces regularly cleaned (e.g., vinyl floor of kitchen and bathroom, kitchen counter and dinner table) were mostly negative.</p> <p>Virus concentration in dust samples from the carpet floor in the bedroom were a least 1 order of magnitude higher than in dust samples from the HVAC filter.</p> | <p>Moderate</p> <p>PREPRINT</p> |

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| <p>Akter, S., Roy, P. C., Ferdaus, A., Ibnat, H., Rubayet UI Alam, A. S. M., Nigar, S., ... Anwar Hossain, M. (2020). Prevalence and stability of SARS CoV-2 RNA on Bangladeshi banknotes. <i>Preprint</i>.</p> | <p>Nov 30, 2020</p> | <p>Prevalence</p> | <p>Prevalence and stability of SARS-CoV-2 on banknotes in circulation.</p> <p>Bangladesh</p> | <p>RT-PCR</p> <p>Prevalence: 425 banknotes collected from 56 entities (shops, restaurants, drivers, etc.) over a period of 3 months in 2 Southern districts of Bangladesh.</p> <p>Stability: banknotes spiked with SARS-CoV-2 (234 samples).</p> | <p>31/425 (7.3%) of banknotes were positive for SARS-CoV-2. The entity with the highest prevalence of positive banknote was a local transport business (14.3%).</p> <p>Stability: N gene detected for up to 72 hours at 35°C. Survival higher on new banknotes compared to older banknotes. New banknotes: more fibrous and compact texture, and more absorbent than old ones.</p> | <p>High</p> <p><i>PREPRINT</i></p> |
| <p>Liu, P., Yang, M., Zhao, X., Guo, Y., Wang, L., Zhang, J., ... Wu, G. (2020). Cold-chain transportation in the frozen food industry may have caused a recurrence of COVID-19 cases in destination: Successful isolation of SARS-CoV-2 virus from the imported frozen cod package surface. <i>Biosafety and Health</i>, 2(4), 199–201.</p> | <p>Nov 19, 2020</p> | <p>Prevalence</p> | <p>Surface samples collected from frozen cod outer packaging, as part of an outbreak investigation of 2 positive COVID-19 cases in workers loading/unloading frozen cod.</p> <p>China</p> | <p>Detection method not specified.</p> <p>421 samples collected.</p> <p>Cleaning of surfaces unlikely prior to sample collection, although not explicitly stated.</p> | <p>50/421 (12%) of samples tested positive for SARS-CoV-2.</p> | <p>Low</p> |

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| <p>Di Carlo, P., Chiacchiarretta, P., Sinjari, B., Aruffo, E., Stuppia, L., De Laurenzi, V., ... Ucciferri, C. (2020). Air and surface measurements of SARS-CoV-2 inside a bus during normal operation. <i>PLoS ONE</i>, 15(11), e0235943. Epub ahead of print.</p> | <p>Nov 5, 2020</p> | <p>Prevalence</p> | <p>Surface and air samples collected from buses.</p> <p>Italy</p> | <p>RT-PCR</p> <p>Samples were collected during the last week of lockdown and the first week after lockdown (May 2020). Air samples were collected every weekday for the length of the bus shift (6.5 hours) and surface samples were collected from 5 commonly touched surfaces (ticket machine and stop buttons) before and after the shift.</p> <p>Cleaning and sanitation performed daily.</p> | <p>None of the samples (surface and air) tested positive.</p> | <p>High</p> |
| <p>Harvey, A. P., Fuhrmeister, E. R., Cantrell, M., Pitol, A. K., Swarthout, J. M., Powers, J. E., ... Pickering, A. J. (2020). Longitudinal monitoring of SARS-CoV-2 RNA on high-touch surfaces in a community setting. <i>Preprint</i>.</p> | <p>Nov 1, 2020</p> | <p>Prevalence</p> | <p>Surface samples from high-touch non-porous surfaces such as handles were collected from community settings.</p> <p>USA</p> | <p>RT-PCR</p> <p>348 samples were collected from 33 unique surfaces (trash can, liquor store, bank, metro door, grocery store, crosswalk buttons, gas station/pump, restaurant, convenience store and post box), throughout a COVID-19 outbreak (Mar 13-Jun 23, 2020).</p> <p>Cleaning frequency/method at sampling locations were not monitored.</p> | <p>29/348 (8.3%) of surface samples were positive for SARS-CoV-2, with 17/33 (52%) of surfaces being positive at least once.</p> <p>The surfaces most frequently contaminated were a trash can handle and a liquor store door handle.</p> <p>The estimated risk of infection from touching a contaminated surface was low (less than 5 in 10,000)</p> | <p>High</p> <p><i>PREPRINT</i></p> |

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| <p>Hu, X., Ni, W., Wang, Z., Ma, G., Pan, B., Dong, L., ... Jiang, F. (2020). The distribution of SARS-CoV-2 contamination on the environmental surfaces during incubation period of covid-19 patients. <i>Ecotoxicology and environmental safety</i>, 208, 111438.</p> | <p>Oct 12, 2020</p> | <p>Prevalence</p> | <p>Hotel</p> <p>3 rooms of quarantined overseas students returned from America who had tested positive for COVID-19 (2 symptomatic and 1 pre-symptomatic)</p> <p>China</p> | <p>RT-PCR</p> <p>Surface samples (light switch, bathroom door knob, toilet, sink, sewer inlet, floor near bed, bedside table, bedding, TV and remote control, telephone and bay window) from within each room were collected within 4 hours of the students testing positive for COVID-19.</p> <p>No disinfectant was used during the hotel quarantine.</p> | <p>Overall, 14 of 41 (34.1%) samples tested positive.</p> <p>46% and 62% of samples from the symptomatic student's rooms tested positive, whereas no samples from the pre-symptomatic student tested positive.</p> <p>All positive samples were from bathroom and bedroom sites (none from living room). The % of positive samples across different surfaces were cotton 60%, ceramic 40%, metal 40%, wood 33% and plastic 16.7%.</p> | <p>Low</p> |
| <p>Luo, L., Liu, D., Zhang, H., Li, Z., Zhen, R., Zhang, X., ... Mao, C. (2020). Air and surface contamination in non-health care settings among 641 environmental specimens of 39 COVID-19 cases. <i>PLoS Neglected Tropical Diseases</i>, 14(10), e0008570.</p> | <p>Oct 9, 2020</p> | <p>Prevalence</p> | <p>Surface samples from sites in which persons with confirmed COVID-19 (9 asymptomatic and 30 symptomatic COVID-19 cases) resided or visited (home or hotel)</p> <p>China</p> | <p>RT-PCR</p> <p>Samples were taken from high touch surfaces and toilets within 3 days of a positive test.</p> | <p>One or more positive surface sample was identified for 9 of the 39 positive cases. Most were within the home or hotel room, particularly within the bathroom (most positive samples being from the floor drain and toilet bowl). No positive samples were found in public areas.</p> | <p>Low</p> |

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| <p>Parker, C. W., Singh, N., Tighe, S., Blachowicz, A., Wood, J. M., Seuylemezian, A., ... Venkateswaran, K. (2020). End-to-end protocol for the detection of SARS-CoV-2 from built environments. <i>mSystems</i>, 5(5), e00771-20.</p> | <p>Oct 6, 2020</p> | <p>Prevalence</p> | <p>Not explicitly stated but from the authors contributions it may be buildings at the NASA Jet Propulsion Laboratory, California Institute of Technology.</p> <p>USA</p> | <p>RT-PCR</p> <p>Environmental samples were collected from 7 different materials found across 10 buildings, in areas with large amounts of pedestrian traffic and deemed high-touch surfaces (stainless steel, Amerstat, plastic, copper, and painted surfaces).</p> <p>Implemented safety practices in the buildings included cleaning, social distancing and mask use.</p> | <p>None of the 368 samples collected tested positive for SARS-CoV-2.</p> | <p>Low</p> |
| <p>Fernández-de-Mera, I. G., Rodriguez Del-Rio, F. J., de la Fuente, J., Perez-Sancho, M., Hervas, D., Moreno, I., ... Gortazar, C. (2020). Detection of environmental SARS-CoV-2 RNA in a high prevalence setting in Spain. <i>Transboundary and Emerging Diseases</i>. Epub ahead of print.</p> | <p>Sep 7, 2020</p> | <p>Prevalence</p> | <p>Surface samples from public service sites in a rural village with ageing population. Household samples (clothing and surfaces) of active and recovered COVID-19 symptomatic cases.</p> <p>Spain</p> | <p>RT-PCR</p> <p>Samples were collected after case numbers had dropped to only 3 active cases in the village at first sampling and none at second sampling (16 days later).</p> <p>Hand and household disinfection were encouraged. Hypochlorite disinfection of public spaces occurred 1-3 times weekly prior/during sampling.</p> | <p>2 of the 6 public service sites surfaces tested positive, the petrol station and the pharmacy.</p> <p>Clothing in both active case households and a surface from 1 of the 6 recovered households tested positive.</p> <p>During repeat sampling clothing from 1 of 7 recovered households tested positive.</p> | <p>Moderate</p> |

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| <p>Piana, A., Colucci, M. E., Valeriani, F., Marcolongo, A., Sotgiu, G., Pasquarella, C., ... Romano Spica, V. (2021). Monitoring COVID-19 transmission risks by RT-PCR tracing of droplets in hospital and living environments. <i>Preprint.</i></p> | <p>Aug 25, 2021</p> | <p>Prevalence</p> | <p>Public buildings (office, fast food outlet, church), outdoor surfaces (handles, handrails, playgrounds) and used handkerchiefs.</p> <p>Italy</p> | <p>RT-PCR</p> <p>Surface samples were collected after the epidemic peak.</p> <p>No information on cleaning protocols of public surfaces.</p> | <p>25 samples from within public buildings (toilets, church pews, floors, wall tiles, phone, computer keyboards, air circulation system) and 16 outdoor samples (handrail, shared scooter grip, bus stop bench, coffee dispenser button, door handle, playground).</p> <p>No SARS-CoV-2 RNA was detectable on any surface sample, although presence of biological fluids were detected.</p> | <p>Moderate</p> <p><i>PREPRINT</i></p> |
| <p>Mouchtouri, V. A., Koureas, M., Kyritsi, M., Vontas, A., ros, Kourentis, L., ... Hadjichristodoulou, C. (2020). Environmental contamination of SARS-CoV-2 on surfaces, air-conditioner and ventilation systems. <i>International Journal of Hygiene and Environmental Health</i>, 230, 113599.</p> | <p>Aug 13, 2020</p> | <p>Prevalence</p> | <p>International ferry boat and nursing home after a COVID-19 outbreak had been identified.</p> <p>Greece</p> | <p>RT-PCR</p> <p>Surface samples were collected once an outbreak was identified and before application of cleaning and disinfection measures.</p> | <p>5 of 9 surface swabs from the ferry boat tested positive (hand contact points such as flour scoop handle, doorknob, bar counter and light switch).</p> <p>4 of 20 surface swabs from the nursing home tested positive (such as patient bed side rail).</p> | <p>Moderate</p> |
| <p>Xie, C., Zhao, H., Li, K., Zhang, Z., Lu, X., Peng, H., ... Lu, J. (2020). The evidence of indirect transmission of SARS-CoV-2 reported in Guangzhou, China. <i>BMC Public Health</i>, 20(1), 1202-1202.</p> | <p>Aug 5, 2020</p> | <p>Prevalence</p> | <p>1 residential building where 2 family clusters of confirmed COVID-19 cases resided.</p> <p>China</p> | <p>RT-PCR</p> <p>21 surface samples from the elevator and houses of 2 families.</p> <p>Before sampling the interior of the elevator had been disinfected several times.</p> | <p>1 of 24 samples tested positive (door handle of first infected family residence) and none of 10 samples tested positive during first and second sampling respectively.</p> <p>It was believed transmission occurred from one family to another via a contaminated elevator button (13 days prior to surface testing).</p> | <p>Low</p> |

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| Jiang, F. C., Jiang, X. L., Wang, Z. G., Meng, Z. H., Shao, S. F., Anderson, B. D., & Ma, M. J. (2020). Detection of severe acute respiratory syndrome coronavirus 2 RNA on surfaces in quarantine rooms . <i>Emerging Infectious Diseases</i> , 26(9), 2162–2164. | May 18, 2020 | Prevalence | Hotel Surface samples collected from 2 rooms in which 2 pre-symptomatic patients had stayed for < 24 hours. China | RT-PCR 11 samples collected on the same surfaces in each room on Mar 20 and 22, 2020. Cleaning of surfaces prior to sample collection unlikely, although not explicitly stated. | 8/22 (36%) of samples were positive for SARS-CoV-2 (55% in 1 room, 18% in the other room). Pillow cover was the only sample positive in both rooms. Duvet cover, sheet, towel, bathroom door handle and light switch were positive in 1 room; faucet was positive in the other room. The other samples (including door handle, toilets and TV remotes) were negative in both rooms. | High |
| Previously reported evidence | | | | | | |
| Wong, J.C.C., Hapuarachichi, H.C., Arivalan, S., Tien, W.P., Koo, C., Mailepessov, D., ... Ng, L.C. (2020). Environmental contamination of SARS-CoV-2 in a non-healthcare setting . <i>International Journal of Environmental Research & Public Health</i> , 18(1), 117. | Dec 26, 2020 | Prevalence | Sites in which persons with confirmed COVID-19 resided or visited. Singapore | RT-PCR Samples collected before and after disinfection of high touch areas (rooms, toilets, elevators) 1-3 days after occupancy. Cleaning and disinfection conducted by professional cleaning companies using various agents. | Two of 428 (0.5%) samples tested positive originating from a bedside wall and bed handle prior to disinfection. Following disinfection and cleaning, repeated collected samples tested negative. | Moderate |

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| <p>Abrahão, J. S., Sacchetto, L., Rezende, I. M., Rodrigues, R. A. L., Crispim, A. P. C., Moura, C., ... Drumond, B. P. (2020). Detection of SARS-CoV-2 RNA on public surfaces in a densely populated urban area of Brazil: A potential tool for monitoring the circulation of infected patients. <i>The Science of the Total Environment</i>, 142645. Epub ahead of print.</p> | <p>Oct 1, 2020</p> | <p>Prevalence</p> | <p>Public places (near hospital and public transportation areas) in region with highest number of reported COVID-19 cases.</p> <p>Brazil</p> | <p>RT-PCR</p> <p>No information on cleaning protocols of public surfaces.</p> | <p>16 of 101 (16.8%) samples tested positive.</p> <p>Positive samples were found on metal and concrete surfaces at hospital bus stations (bench, ground), hospital sidewalks, bus terminals (handrails), and public square seating (table and benches).</p> | <p>High</p> |
| <p>Marshall, D., Bois, F., Jensen, S. K. S., Linde, S. A., Higby, R., Remy-McCort, Y., ... Martin, G.G. (2020). Sentinel coronavirus environmental monitoring can contribute to detecting asymptomatic SARS-CoV-2 virus spreaders and can verify effectiveness of workplace COVID-19 controls. <i>Microbial Risk Analysis</i>. Epub ahead of print.</p> | <p>Aug 30, 2020</p> | <p>Prevalence</p> | <p>Workplace</p> <p>9 office and mixed-used industrial locations.</p> <p>Europe USA</p> | <p>RT-PCR</p> <p>Samples were collected near end of work shifts and prior to disinfection and cleaning.</p> | <p>Number or percent of positive samples not reported.</p> <p>Workplaces with positive samples were more likely to have employees with confirmed cases of COVID-19.</p> <p>Highest positive sample rates were found among door handles and shared furniture (break room chairs, workbenches).</p> | <p>Moderate</p> |

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| Yamagishi, T., Ohnishi, M., Matsunaga, N., Kakimoto, K., Kamiya, H., Okamoto, K., ... Wakita, T. (2020). Environmental sampling for severe acute respiratory syndrome coronavirus 2 during COVID-19 outbreak in the Diamond Princess cruise ship . <i>The Journal of Infectious Diseases</i> , 222(7), 1098-1102. | Jul 21, 2020 | Prevalence | Cruise ship Surface samples from vacant cabins of those with confirmed COVID-19 cases, cabins with no confirmed cases, and common areas. | Reverse transcription polymerase chain reaction (RT-PCR) used to detect presence of SARS-CoV-2 RNA. Samples collected 7-9 days after disinfection with 5% hydrogen peroxide. | No viable virus was detected in any of the samples. 58 of 601 (10%) samples tested positive from cabins with confirmed COVID-19 cases 1-17 days after the cabins were vacated, but not from non-case-cabins. The virus was most often detected on the bathroom floor near the toilet and bed pillows. | High |
| Döhla, M., Wilbring, G., Schulte, B., Kümmerer, B.M., Diegmann, C., Sib, E., ... Schmithausen, R.M. (2020). SARS-CoV-2 in environmental samples of quarantined households . <i>Preprint</i> . | Jun 2, 2020 | Prevalence | Households under quarantine with at least one confirmed COVID-19 case. Germany | RT-PCR Viral culture test Surface samples collected from frequently shared objects (e.g., door handles, remote control). | 4 of 119 (3.36%) samples tested positive. Positive samples were found on electronic devices, knobs/handles, and furniture. Viral culturing detected no viable virus from the samples. | Moderate PREPRINT |
| Bloise, I., Gómez-Arroyo, B., & García-Rodríguez, J. (2020). Detection of SARS-CoV-2 on high-touch surfaces in a clinical microbiology laboratory . <i>The Journal of Hospital Infection</i> , 105(4), 784-786. | May 15, 2020 | Prevalence | Clinical microbiology laboratory with high density of samples tested for COVID-19. Samples from high touch surfaces. Spain | RT-PCR Cleaning and disinfection protocols not reported. | 4 of 22 (18%) samples tested positive. Positive samples were found on commonly used objects, such as keyboards, telephones and computer mouse, representing potential sources of infection for laboratory personnel. | Moderate |

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| Lee, S.E., Lee, D.Y., Lee, W.G., Kang, B., Jang, Y.S., Ryu, B., ... Lee, E. (2020). Detection of novel coronavirus on the surface of environmental materials contaminated by COVID-19 patients in the Republic of Korea. <i>Osong Public Health and Research Perspectives</i> , 11(3), 128–132. | May 8, 2020 | Prevalence | A rehabilitation centre and an apartment building complex with COVID-19 outbreaks. Korea | RT-PCR Samples collected from high touch surfaces (e.g. door handles). | 2 of 12 (16.7%) samples from communal facilities where disinfection and cleaning had not been conducted prior to collection tested positive. Both samples were from a door handle of a COVID-19 positive patient's room. | Moderate |
| Laboratory Studies | | | | | | |
| New evidence reported March 5, 2021 | | | | | | |
| Morris, D., Yinda, K. C., Gamble, A., Rossine, F. W., Huang, Q., Bushmaker, T., ... Lloyd-Smith, J.O. (2020). Mechanistic theory predicts the effects of temperature and humidity on inactivation of SARS-CoV-2 and other enveloped viruses. <i>Preprint</i> . | Nov 13, 2020 | Laboratory | SARS-CoV-2 suspension deposited onto polypropylene plastic surface at different temperatures and humidities. USA | Viral culture test | Viral decay was faster with increasing temperature with decay at 27°C 5-10 X faster than decay at 10°C. Virus decay was fastest at 65% relative humidity and tended to be slower at lower (40%) or higher (85%) relative humidities. | Not appraised <i>PREPRINT</i> |

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| <p>Harbourt, D. E., Haddow, A. D., Piper, A. E., Bloomfield, H., Kearney, B. J., Fetterer, D., ... Minogue, T. (2020). Modeling the stability of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) on skin, currency, and clothing. <i>PLoS Neglected Tropical Diseases</i>, 14(11), e0008831. Epub ahead of print.</p> | <p>Nov 9, 2020</p> | <p>Laboratory</p> | <p>Evaluated surface stability of SARS-CoV-2 on swine skin, banknotes and scrub fabric at different temperatures.</p> <p>USA</p> | <p>Viral culture test</p> | <p>SARS-CoV-2 remained stable in swine skin for the full experiment of 14 days at 4°C; 96 hours at 22°C and up to 8 hours at 37°C.</p> <p>At 4°C, SARS-CoV-2 remained quantifiable on bank notes and clothing for up to 96 hours.</p> <p>At 22°C, SARS-CoV-2 decay was quicker, being negative after 24 hours on bank notes and 8 hours on clothing.</p> <p>At 37°C SARS-CoV-2 was not detected after 4 hours on clothing and 8 hours on bank notes.</p> | <p>Not appraised</p> |
| <p>Magurano, F., Baggieri, M., Marchi, A., Rezza, G., & Nicoletti, L. (2020). SARS-CoV-2 infection: the environmental endurance of the virus can be influenced by the increase of temperature. <i>Clinical Microbiology and Infection</i>. Epub ahead of print.</p> | <p>Nov 5, 2020</p> | <p>Laboratory</p> | <p>SARS-CoV-2 suspension deposited onto polypropylene plastic at different temperatures and harvested at different predefined time-points (up to 7 days) at a relative humidity of 35-45%.</p> <p>Italy</p> | <p>Viral culture test</p> | <p>At 20-25°C, virus infectivity on a plastic surface reduced rapidly in the first 24-36 hours. At 28°C, same reduction was observed in the first 8-12 hours.</p> <p>At both 20-25°C and at 28°C, SARS-CoV-2 maintain its ability to infect cells on a plastic surface for up to 84 hours but was not detectable at 96 hours.</p> | <p>Not appraised</p> |

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| <p>Riddell, S., Goldie, S., Hill, A., Eagles, D., & Drew, T.W. (2020). The effect of temperature on persistence of SARS-CoV-2 on common surfaces. <i>Virology Journal</i> (17), 145.</p> | <p>Oct 7, 2020</p> | <p>Laboratory</p> | <p>SARS-CoV-2 suspension dried onto glass, stainless steel, paper and polymer bank notes, vinyl, cloth in the dark for up to 28 days at 20°C, 30°C and 40°C and 50% humidity.</p> <p>Australia</p> | <p>Viral culture test</p> | <p>Infectious virus was still detectable after 28 days at 20°C, for all non-porous surfaces tested. For porous (cloth) no infectious virus was detected past day 14, with the majority absorbed soon after application.</p> <p>At 30°C infectious virus was detectable for 7 days for most non-porous surfaces and 3 days for cloth.</p> <p>At 40°C infectious virus was not recovered past 48 hours for non-porous and 24 hours for cloth.</p> | <p>Not appraised</p> |
| <p>Matson, M.J., Yinda, C.K., Seifert, S.N., Bushmaker, T., Fischer, R.J., van Doremalen, N., ... Munster, J.J. (2020). Effect of environmental conditions on SARS-CoV-2 stability in human nasal mucus and sputum. <i>Emerging Infectious Diseases</i>, 26(9), 2276-2278.</p> | <p>Jun 8, 2020</p> | <p>Laboratory</p> | <p>SARS-CoV-2 mixed with commercially available nasal mucus and sputum and deposited on polypropylene discs at different temperatures and humidities.</p> <p>USA</p> | <p>Viral culture test and RT-PCR</p> | <p>In surface nasal mucus, half-life of SARS-CoV-2 was shorter at 27°C/85%RH than 21°C/40%RH (p=0.002) and 4°C/40%RH (p=0.0007). Half-life was shorter on the surface compared to in liquid nasal mucus (p=0.01).</p> <p>In surface sputum, half-life of SARS-CoV-2 was shorter at 27°C/85%RH (p=0.0002) and 21°C/40%RH (p=0.004) compared with 4°C/40%RH, with no difference in half-life between liquid and surface sputum.</p> <p>Infectious virus persisted in both nasal mucus and sputum on surfaces for ~24 hours and SARS-CoV-2 RNA remained detectable for ≥7 days in all surface samples.</p> | <p>Not appraised</p> |

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| <p>Kratze, A., Steiner, S., Todt, D., V'kovski, P., Brueggemann, Y., Steinmann, J. ... Pfaender, S. (2020). Temperature-dependent surface stability of SARS-CoV-2. <i>Journal of Infection</i> 81(3), 452-482.</p> | <p>Jun 2, 2020</p> | <p>Laboratory</p> | <p>SARS-CoV-2 suspensions dried onto metal discs at 4°C, RT and 30°C for up to 9 days at 30-40% humidity.</p> <p>Germany</p> | <p>Viral culture test</p> | <p>Drying reduced infectivity of virus by 100-fold, but then remained stable for 4-8 hours. Beyond 8 hours there was a stable, slow decline of viral titres at all temperatures over several days.</p> <p>Detectable infectious virus was still present after 180 hours. The median half-life is predicted to be 9.1 hours at RT, 12.9 hours at 4°C, and 17.9 hours at 30°.</p> <p>Surface stability may be due to constant humidity.</p> | <p>Not appraised</p> |
| <p>Chin, A. W. H., Chu, J. T. S., Perera, M. R. A., Hui, K. P. Y., Yen, H.-L., Chan, M. C. W., . . . Poon, L. L. M. (2020). Stability of SARS-CoV-2 in different environmental conditions. <i>The Lancet. Microbe</i>, 1(1), e10. Epub ahead of print.</p> | <p>Apr 2, 2020</p> | <p>Laboratory</p> | <p>SARS-CoV-2 added to a surface (printing and tissue paper, treated wood, cloth, glass, banknote, stainless steel, plastic, surgical mask) at different temperatures and pHs and added to various disinfectants for up to 14 days.</p> <p>China</p> | <p>Viral culture test</p> | <p>After 3 hours, no infectious virus could be recovered from printing and tissue paper, after 2 days on treated wood and cloth, 4 days on glass and banknote or 7 days on stainless steel or plastic. A detectable level of infectious virus was present on the outer layer of a surgical mask on day 7.</p> <p>SARS-CoV-2 is extremely stable in a wide range of pH values at room temperature. It is highly stable at 4°C, but at 70°C the time for virus inactivation was reduced to 5 mins.</p> <p>Except for hand soap, no infectious virus could be detected after a 5-minute incubation at room temperature.</p> | <p>Not appraised</p> |

| Previously reported evidence | | | | | | |
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| <p>Liu, Y., Li, T., Deng, Y., Liu, S., Zhang, D., Li, H., ... Li, J. (2020). Stability of SARS-CoV-2 on environmental surfaces and in human excreta. <i>The Journal of Hospital Infection</i>. Epub ahead of print.</p> | <p>Oct 30, 2020</p> | <p>Laboratory</p> | <p>Steel, plastic, glass, ceramics, paper, cotton, wood, latex gloves, surgical mask deposited and left for 7 days.</p> <p>China</p> | <p>Viral culture test</p> | <p>The virus remained stable and viable for seven days on surfaces of plastic, stainless steel, glass, ceramics, wood, latex gloves and surgical mask.</p> <p>The virus did not remain infectious after 4 days on cotton clothes and after 5 days on paper. In both of these materials, rapid loss of virus infectivity was observed within 1 hour after incubation.</p> <p>Across most of the tested conditions, in the initial phase of viral decay, loss of infectivity was rapid, whereas in the terminal phase, viral infectivity decreased slowly.</p> | <p>Not appraised</p> |
| <p>Pelisser, M., Thompson, J., Majra, D., Youhanna, S., Stebbing, J., & Davies, P. (2020). Sports balls as potential SARS-CoV-2 transmission vectors. <i>Public Health in Practice</i> 1, 100029. Epub ahead of print.</p> | <p>Jul 10, 2020</p> | <p>Laboratory</p> | <p>Sports equipment with inactivated virus pipetted directly onto the surface in a lab</p> <p>UK</p> | <p>Testing protocol not reported</p> | <p>Surfaces of sports balls were tested before and after disinfection, and after use on a grass field. All samples tested negative.</p> <p>The authors note a limitation to the study may have been the method used to transfer the virus to surfaces using polyester swabs which may not have been effective.</p> | <p>Not appraised</p> |

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| <p>Pastorino, B., Touret, F., Gilles, M., de Lamballerie, X., & Charrel, R.N. (2020). Prolonged infectivity of SARS-CoV-2 in fomites. <i>Emerging Infectious Diseases</i>. 26(9), 2256-2257.</p> | <p>Jun 24, 2020</p> | <p>Laboratory</p> | <p>SARS-CoV-2 deposited on polystyrene plastic, aluminum, and glass for 96 hours</p> <p>France</p> | <p>Viral culture test</p> | <p>SARS-CoV-2 demonstrated viral stability for 96 hours on all tested surfaces.</p> <p>Protein mediums increased SARS-CoV-2 infectivity, suggesting that protein-rich mediums such as airway secretions can protect the expelled virus, potentially enhancing persistence and transmission via contaminated surfaces.</p> | <p>Not appraised</p> |
|--|---------------------|-------------------|--|---------------------------|---|----------------------|

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